Il Farmaco 53 (1998) 519-524

#### Short Communication

# Pyrazole analogues of prazosin

# Giuseppe Ermondi, Donatella Boschi, Antonella Di Stilo, Carla Tironi, Alberto Gasco\*

Dipartimento di Scienza e Tecnologia del Farmaco, Università di Torino, Via P. Giuria 9, 10125 Turin, Italy

Received 12 January 1998; accepted 9 July 1998

#### Abstract

A series of analogues of prazosin, in which 1-methyl or 1-phenylpyrazole moieties were substituted for the furan ring, were synthesized and studied for their  $\alpha_1$ -adrenoceptor antagonist activity. The role of the five member heterocyclic substructures in determining the affinity for the  $\alpha_1$ -receptor is briefly discussed. © 1998 Elsevier Science S.A. All rights reserved.

Keywords: α<sub>1</sub>-Adrenoceptor antagonist; Prazosin analogues; Pyrazole derivatives; Molecular modelling

#### 1. Introduction

Structure-activity relationships (SARs) in prazosin 1, a well-known α<sub>1</sub>-adrenoceptor antagonist, have been the object of several studies. The role of the 2,4-diamino-6,7-dimethoxyguinazoline moiety has been established [1-3]. This substructure, at physiological pH, is partially protonated at the endocyclic N-1 nitrogen and acts as a conformationally restricted form for the noradrenaline cation. The role of the piperazine ring has been analysed [4] and conformational analysis, molecular modelling, and quantitative structureactivity relationship studies have been carried out on sets of 2,4-diamino-6,7-dimethoxy-2-substituted quinazolines [5,6]. The substitution of a number of aromatic five member heterocyclic rings for furan in 1 has been explored, principally in patent literature. To our knowledge only one paper, concerning the study of a number of isoxazole analogues of prazosin, has been devoted to this subject [7]. In addition, in a recent work we reported the replacement of R-substituted 1,2,5-oxadiazoles N-oxides (furoxans) for the furan ring to design hybrid molecules able to display simultaneously  $\alpha_1$ adrenoceptor antagonistic activity and NO mediated vasodilating properties [8,9]. In that investigation, the corresponding 1,2,5-oxadiazole analogues (furazans) were also considered. In this paper, we report synthesis,  $\alpha_1$ -antagonist properties and SARs of pyrazole congeners of prazosin, belonging to series I, II and III (Scheme 1).

III: 5a, R=CH3; 5b, R=C6H5

Scheme 1. Prazosin (1) and its pyrazole congeners (series I, II, III).

<sup>\*</sup> Corresponding author.

Scheme 2. Preparation of compounds 3a-5a and 3b-5b.

Table 1  $^{1}$ H NMR chemical shifts  $\delta$  and coupling constants J of the compounds 3a,b, 4a,b and 5a,b

Hydrogen	3-Substituted pyrazoles		4-Substituted pyrazoles		5-Substituted pyrazoles	
	$3a (R = CH_3)$	<b>3b</b> $(R = C_6H_5)$	$4a (R = CH_3)$	<b>4b</b> $(R = C_6H_5)$	$5a (R = CH_3)$	<b>5b</b> $(R = C_6H_5)$
5	7.46 (s, 1H)	7.46 (s, 1H)	7.44 (s, 1H)	7.48 (s, 1H)	7.46 (s, 1H)	7.45 (s, 1H)
8	6.78 (s, 1H)	6.77 (s, 1H)	6.76 (s, 1H)	6.80 (s, 1H)	6.78 (s, 1H)	6.74 (s, 1H) b
6-OCH <sub>3</sub>	3.81 (s, 3H)	3.81 (s, 3H)	3.81 (s, 3H)	3.81 (s, 3H)	3.81 (s, 3H)	3.80 (s, 3H)
7-OCH <sub>3</sub>	3.85 (s, 3H)	3.85 (s, 3H)	3.85 (s, 3H)	3.86 (s, 3H)	3.85 (s, 3H)	3.84 (s, 3H)
N	4.0-3.7 (m, 8H)	4.0–3.7 (m, 8H)	4.0–3.7 (m, 8H)	4.0–3.7 (m, 8H)	4.0–3.5 (m, 8H)	3.7–3.2 (m, 8H)
3'			7.72 (s, 1H)	8.05 (s, 1H)	7.51 (d, $J = 1.9$ Hz, 1H)	7.83 (d, $J = 1.7$ Hz, 1H)
4'	6.59 (d, $J = 2.2$ Hz, 1H)	6.88 (d, $J = 2.5$ Hz, 1H)			6.51 (d, $J = 1.9$ Hz, 1H)	6.74 (d, $J = 1.7$ Hz, 1H) <sup>b</sup>
5'	7.79 (d, $J = 2.2 \text{ Hz}$ , 1H)	8.62 (d, $J = 2.5$ Hz, 1H)	8.11 (s, 1H)	8.88 (s, 1H)		
N-CH <sub>3</sub>	3.92 (s, 3H)		3.88 (s, 3H)		3.88 (s, 3H)	
N-C <sub>6</sub> H <sub>5</sub>	,	7.3-8.0 (m, 5H)	•	7.3-8.0 (m, 5H)	•	7.4-7.6 (m, 5H)
NH <sub>2</sub>	7.24 (br, 2H)	7.18 (br, 2H)	7.18 (br, 2H)	7.26 (br, 2H)	7.2 (br, 2H)	7.19 (br, 2H)

<sup>&</sup>lt;sup>a</sup>  $\delta$  in ppm from TMS. Solvent DMSO-d<sub>6</sub>.

In these compounds the quinazolinylpiperazinocarbonyl moiety is linked at positions 3, 4 and 5, respectively, of the pyrazole ring. This structural feature, combined with phenyl or methyl substitution at N-1 of the heterocyclic system, modulates the shape and volume of this molecular region and renders the three series suitable probes for exploring the receptor area fitted by the pyrazole ring.

## 2. Experimental

#### 2.1. Chemistry

Melting points were measured on a Büchi 530 capillary apparatus and are uncorrected. The compounds were routinely checked by infrared spectrometry (Shimadzu FT-IR

<sup>&</sup>lt;sup>b</sup> Overlapping signals.

Table 2  $^{13}$ C NMR chemical shifts  $\delta$  \* of the compounds 3a,b, 4a,b and 5a,b

Carbon	3-Substituted pyrazoles		4-Substituted pyrazoles		5-Substituted pyrazoles	
	$3a (R = CH_3)$	3b $(R = C_6H_5)$	$4a (R = CH_3)$	<b>4b</b> $(R = C_6H_5)$	$5a (R = CH_3)$	<b>5b</b> $(R = C_6H_5)$
2	158.1	158.4	158.4	158.1	158.3	158.0
4	161.2	161.2	161.2	161.2	161.3	161.2
4a	103.0	103.1	103.0	103.0	103.1	103.0
5	103.8	103.7	103.7	103.8	103.7	103.7
6	145.2	145.1	145.1	145.2	145.2	145.2
7	154.3	154.3	154.3	154.4	154.4	154.3
8	105.1	105.3	105.3	105.0	105.3	105.2
8a	148.4	148.9	148.8	148.3	148.8	148.5
6-OCH <sub>3</sub>	55.9	55.9	55.9	55.9	55.9	55.9
7-OCH <sub>3</sub>	55.5	55.5	55.5	55.5	55.5	55.5
	46.5-44.3, 43.7-	46.7-44.3, 43.6-42.2	43.8	47.0 (vbr), 43.8	47.0-43.5, 41.8	46.5-43.3, 43.0-41.6
	42.1			(br)		·
C=O	162.0	161.7	162.8	162.5	160.6	160.7
3'	144.2	148.0	139.3	141.4	137.5	140.3
4'	106.2	110.1	116.5	116.8	106.7	108.3
5'	131.7	128.8	132.6	129.1	135.4	136.2
CH <sub>3</sub>	38.7		38.7		37.8	
C <sub>6</sub> H <sub>5</sub> -C <sub>i</sub>		139.4		139.3		139.4
C <sub>6</sub> H <sub>5</sub> -C <sub>o</sub>		118.9		116.9		123.0
C <sub>6</sub> H <sub>5</sub> -C <sub>m</sub>		129.7		129.6		129.5
C <sub>6</sub> H <sub>5</sub> -C <sub>p</sub>		127.0		127.0		127.9

<sup>&</sup>lt;sup>a</sup>  $\delta$  in ppm from TMS. Solvent DMSO-d<sub>6</sub>.

Fig. 1. Torsion angles considered in the conformational analysis.

8101 M),  $^1$ H and  $^{13}$ C nuclear magnetic resonance at 200 and 50 MHz, respectively (Bruker AC-200 spectrometer), and mass spectrometry (Finnigan-Mat TSQ-700 spectrometer). Thin layer chromatography (TLC) was carried out on  $5 \times 20$  cm plates precoated with Merck silica gel  $60\,\mathrm{F}_{254}$ , with a layer thickness of 0.25 mm. Anhydrous magnesium sulfate was used as the drying agent of the organic extracts. Solvent removal was achieved under reduced pressure at room temperature. Elemental analyses of the new compounds were performed by REDOX (Cologno M.) and the results are within  $\pm 0.4\%$  of theoretical values.

The carboxylic acid 2a [10], 2b [11], 2c [12], 2d [13], 2e [14], 2f [15], and the intermediate 6 [16] were synthesized according to literature methods.

# 2.2. General procedure for the preparation of compounds 3a,b, 4a,b and 5a,b

These compounds were obtained according to a modified procedure of the method reported in Ref. [8]. A solution of the appropriate pyrazolcarboxylic acid **2a-f** (2 mmol) in thionyl chloride (8 ml) containing a few drops of anhydrous dimethylformamide was heated at reflux for three hours. The excess of thionyl chloride was removed in vacuo and the residue, dissolved in dry tetrahydrofuran (3 ml), was added dropwise to a stirred solution of 4-amino-6,7-dimethoxy-2-piperazinoquinazoline **6** (0.58 g, 2 mmol) dissolved in a mixture of water/methanol (1:10) (15 ml) containing potassium bicarbonate (0.3 g, 3 mmol). The reaction mixture was stirred at 20°C for 2 h then the precipitate was filtered and recrystallized from the appropriate solvent to give products as free bases.

**3a**: 1-(4-amino-6,7-dimethoxy-2-quinazolyl)-4-(1-methyl-3-pyrazolylcarbonyl) piperazine. Methanol was used as cosolvent in the reaction instead of water/methanol. Yield 50%, m.p.  $215-216^{\circ}$ C (from EtOAc). *Anal.* ( $C_{19}H_{23}N_7O_3$ ) C, H, N.

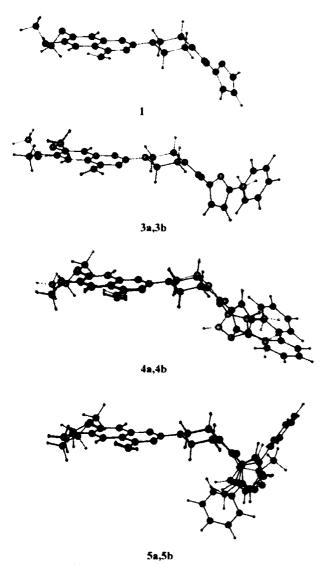


Fig. 2. Side view of the stable conformers.

**4a**: 1-(4-amino-6,7-dimethoxy-2-quinazolyl)-4-(1-methyl-4-pyrazolylcarbonyl)piperazine. Yield 78%, m.p.  $254-255^{\circ}$ C (from MeOH). *Anal*. ( $C_{19}H_{23}N_7O_3 \cdot CH_3OH$ ) C, H, N.

**5a**: 1-(4-amino-6,7-dimethoxy-2-quinazolyl)-4-(1-methyl-5-pyrazolylcarbonyl) piperazine. Methanol was used as cosolvent in the reaction instead of water/methanol. Yield 83%, m.p. 228–229°C (from MeOH). *Anal.*  $(C_{19}H_{23}N_7O_3\cdot 0.5\ CH_3OH)\ C, H, N.$ 

**3b**: 1-(4-amino-6,7-dimethoxy-2-quinazolyl)-4-(1-phenyl-3-pyrazolylcarbonyl) piperazine. KHCO<sub>3</sub> (0.6 g, 6 mmol) was used in the reaction. Yield 93%, m.p. 256–257°C (from MeOH). *Anal*. ( $C_{24}H_{25}N_7O_3$ ) C, H, N.

**4b**: 1-(4-amino-6,7-dimethoxy-2-quinazolyl)-4-(1-phenyl-4-pyrazolylcarbonyl) piperazine. Yield 82%, m.p. 230–231°C (from MeOH). Anal. ( $C_{24}H_{25}N_7O_3 \cdot CH_3OH$ ) C, H, N.

**5b**: 1-(4-amino-6,7-dimethoxy-2-quinazolyl)-4-(1-phenyl-5-pyrazolylcarbonyl)piperazine. Yield 93%, m.p.

244–247°C (from dioxane). *Anal.* ( $C_{24}H_{25}N_7O_3 \cdot 0.5 H_2O$ ) C, H, N.

#### 2.3. Pharmacology

Thoracic aorta were isolated from male Wistar rats (200-250 g) anesthetized with CO<sub>2</sub> and sacrificed by decapitation. All animals were dealt with in a humane way in accordance with recognized guidelines on animal experimentation. The vessels were helically cut, the endothelium removed and two strips were obtained from each aorta. The tissues were suspended under a tension of 1 g in organ baths containing 30 ml of Krebs-Heinselet solution of the following composition (mM): NaCl, 137; KCl, 2.7; MgCl<sub>2</sub>, 0.5; CaCl<sub>2</sub>, 1.4; NaH<sub>2</sub>PO<sub>4</sub>, 0.5; NaHCO<sub>3</sub>, 8.9; glucose, 8.3; ascorbic acid, 0.1. Desmethylimipramine hydrochloride ( $1 \times 10^{-7}$  M), deoxycorticosterone acetate  $(5 \times 10^{-6} \text{ M})$ , propranol  $(5 \times 10^{-6} \text{ M})$ M) and yohimbine hydrochloride  $(1 \times 10^{-7} \text{ M})$  were added to the solution to prevent neuronal and extraneuronal uptake of ( – )noradrenaline and to block  $\beta$ - and  $\alpha_2$ -adrenoceptors, respectively. The medium was maintained at 37°C and at pH 7.4 (by gassing with  $95\%O_2-5\%CO_2$ ). The aortic strips were allowed to equilibrate for 2 h before starting the experiments. ( – )Noradrenaline induced contractions were determined cumulatively in the absence or presence of the antagonist, which was pre-incubated for 30 minutes. One of the two strips cut from each aorta served as a control while a dose-response curve in the presence of the antagonist was performed on the other strip.

### 3. Results and discussion

The compounds 3a–5a and 3b–5b were prepared according to the reaction sequence reported in Scheme 2. The appropriate carboxylic acid, synthesized according to literature methods, was treated with boiling thionyl chloride and the crude chloride obtained was conjugated with 6 in THF solution, in the presence of KHCO<sub>3</sub>. The structures of the final products were confirmed by <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy. The chemical shifts are reported in Tables 1 and 2.

In addition to spectral characterization, a systematic search for accessible (minimum energy) conformations of all the compounds was carried out according to the approaches reported in Refs. [5,17]. Initial geometries of the compounds 3a-5a, 3b-5b and 1 were built by the standard libraries of the Molecular Simulation BUILDER module. The conformations in the prazosin analogues are described by three torsion angles, as shown in Fig. 1. The torsion angle  $\omega_a$  was found to be the same for all compounds  $(20^{\circ}\pm2^{\circ})$ . The torsion angles  $\omega_1$  and  $\omega_2$  were systematically modified by rigid rotations in steps of 15°. At each step, energy was calculated by using the molecular mechanics methods. The conformations with energy greater than 500 kcal mol<sup>-1</sup> were eliminated in order to reduce CPU times. The remaining conformers were optimized by using the molecular mechan-

Table 3  $\alpha^1$ -Adrenoceptor antagonistic activities and AM1 energies of the stable conformers of the compounds 3a,b, 4a,b and 5a,b

Comp.	pA <sub>2</sub> ± 95%CL <sup>a</sup>	Slope ± 95%CL <sup>a</sup>	Conformer	Angle $\omega_2^b$ (deg)	$\Delta E^{c}$ (kcal mol <sup>-1</sup> )
3a	8.68 ± 0.16	$1.07 \pm 0.14$	1	-117.6	0.0
3b	$8.76 \pm 0.12$	$1.00\pm0.16$	1	-117.8	0.0
4a	$8.93 \pm 0.17$	$0.98 \pm 0.14$	1	-141.1	0.0
			2	-46.0	1.8
4b	$8.26 \pm 0.17$	$1.01 \pm 0.17$	1	-140.0	0.0
			2	-47.3	1.8
5a	$8.55 \pm 0.17$	$0.99 \pm 0.18$	1	- 104.8	0.0
	<del></del>		2	75.0	0.3
5b	$8.20 \pm 0.18$	$1.08 \pm 0.16$	1	-82.3	0.7
	_	<del>-</del>	2	57.7	0.0
1	$9.81 \pm 0.27$	$0.89 \pm 0.15$	1	- 144.6	0.0

<sup>&</sup>lt;sup>a</sup> CL = confidence limits.

<sup>&</sup>lt;sup>b</sup> Torsion angle defined by the following atoms:



Atom 4 is represented by N in series I and III and by the C adjacent to N-R in series II.

 $^{\circ} \Delta E$  is the energy difference between the conformer and the minimum.

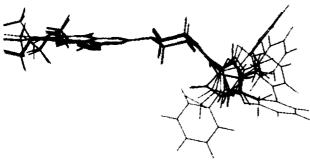


Fig. 3. Total view of the superimposed stable conformers. The superimposition was made using the common quinazolinylpiperazinocarbonyl moiety as the base.

ics methods and the identical geometries were discarded (r.m.s. < 0.3). Finally, the conformers were optimized again by using the AM1 semiempirical approximation. All calculations were performed in vacuo.

The following modules of the molecular modelling software produced by Molecular Simulations Inc. (San Diego, CA) were used: INSIGHT II 95.0, BUILDER 95.0, DISCOVER 95.0/3.00. The CFF91 force field [18] was used in the molecular mechanics calculations. The semiempirical calculations were performed by AM1 [19] Hamiltonian with SPARTAN software (Wavefunction Inc., Irvine, CA). All the software ran on a R4400 Silicon Graphics workstation. Fig. 2 shows the side view of the minimum energy conformations for the compounds 1, 3a–5a and 3b–5b, chosen within 3 kcal mol<sup>-1</sup> of their absolute minima. Models 1 and 3a,b were able to adopt only the absolute minimum conformation, whereas each member of series II and III shows two minimum conformations ( $\Delta E < 2$  kcal mol<sup>-1</sup>) in which  $\omega_2$ 

differs by about 180° (Table 3). Fig. 3 shows the side view of the supermolecule obtained by superimposing the accessible conformations of all the compounds, prazosin included. The rigid superimposition of the 10 minimum conformations and prazosin was made using the common portion of the molecules, the quinazolinylpiperazinocarbonyl moiety, as the base.

The  $\alpha_1$ -Adrenoceptor blocking activity of the products **3a**–**5a**, **3b**–**5b** was assessed by antagonism of ( – ) noradrenaline induced contractions on rat aortic strips. It is known that  $\alpha_1$ -adrenoceptor can exist in three subtypes designated  $\alpha_{1A}$ ,  $\alpha_{1B}$  and  $\alpha_{1D}$ , respectively. On rat aorta strips the  $\alpha_{1D}$  subtype is principally present [20]. The antagonist potency of the compounds, expressed as pA<sub>2</sub>, is reported in Table 3.  $\alpha_2$ -Adrenoceptor blocking activities were tested by antagonism of clonidine induced depression of the twitch responses of a field stimulated prostatic portion of rat vas deferens. None of the compounds displayed any  $\alpha_2$ -antagonist activity at  $10^{-5}$  M concentration.

Analysis of Table 3 shows that all of the members of the series display lower affinity for  $\alpha_1$ -receptor than prazosin. The compounds behave in a similar way, indeed their pA<sub>2</sub> values range between 8.20 (**5b**) and 8.93 (**4a**). A possible explanation for the close receptor affinities of methyl and phenyl analogues in each of the three series is that N-1 pyrazole substituents are projected into a freely accessible space, without giving relevant interactions. In this case, since conformational analysis shows that the pyrazole system of the compounds **4a** and **4b** occupies the same area fitted by the furan of prazosin (Fig. 3), we could infer that the different antagonism displayed by these substances with respect to the

lead is principally due to the different electronic and lipophilic properties of the two heterocyclic moieties. By way of contrast, in the derivatives of the series I and III, in which  $\omega_2$  angles are rather different from those of prazosin (Table 3), conformational effects could also be involved in the modulation of the antagonism.

#### References

- S.F. Campbell, α-Adrenoreceptor modulation. In: A.S. Horn, C.F. De Rauter (Eds.), X-ray Crystallography and Drug Action, Clarendon, Oxford, 1984, pp. 347–366.
- [2] P.G. De Benedetti, M.C. Menziani, G. Rastelli, M. Cocchi, Molecular orbital study of the nitrogen basicity of prazosin analogues in relation to their α<sub>1</sub>-adrenoceptor binding affinity, J. Mol. Struct. (Theochem.) 233 (1991) 343–351.
- [3] D. Boschi, A. Di Stilo, R. Fruttero, C. Medana, G. Sorba, A. Gasco, α<sub>1</sub>-Adrenoreceptor blocking activity of some ring-open analogues of prazosin, Arch. Pharm. (Weinheim) 327 (1994) 661-667.
- [4] D. Giardinà, U. Gulini, M. Massi, M.G. Piloni, P. Pompei, G. Regaiani, C. Melchiorre, Structure-activity relationships in prazosin-related compounds. 2. Role of the piperazine ring on α-blocking activity, J. Med. Chem. 36 (1993) 690-698.
- [5] G. Rastelli, F. Fanelli, M.C. Menziani, M. Cocchi, P.G. De Benedetti, Conformational analysis, molecular modeling and quantitative structure-activity relationship studies of 2,4-diamino-6,7-dimethoxy-1substituted quinazoline α<sub>1</sub>-adrenergic antagonists, J. Mol. Struct. (Theochem.) 251 (1991) 308-318.
- [6] M. Cocchi, G. Cruciani, M.C. Menziani, P.G. De Benedetti, Use of advanced chemometric tools and comparison of different 3D descriptors in QSAR analysis of prazosin analog α<sub>1</sub>-adrenergic antagonists. In: C.G. Wermuth (Ed.), Trends in QSAR and Molecular Modeling 92, Escom, Leiden, 1993, pp. 527–529.
- [7] A. Carenzi, D. Chiarino, M. Napoletano, A. Reggiani, R. Sala, New

- isoxazole derivatives provided with antihypertensive activity, Arzneim. Forsch. Drug Res. 39 (1989) 642-646.
- [8] A. Di Stilo, R. Fruttero, D. Boschi, A.M. Gasco, G. Sorba, A. Gasco, M. Orsetti, Use of nitric oxide releasing furoxan system in the design of 'hybrids'; substitution of furoxan moieties for the furan ring in prazosin, Med. Chem. Res. 3 (1993) 554-566.
- [9] R. Fruttero, D. Boschi, A. Di Stilo, A. Gasco, The furoxan system as a useful tool for balancing hybrids with mixed α<sub>1</sub>-antagonist and NOlike vasodilator activities, J. Med. Chem. 38 (1995) 4944–4949.
- [10] C. Wijnberger, C.L. Habraken, The electron-releasing capacity of the pyrazole ring, J. Heterocycl. Chem. 6 (1969) 545-551.
- [11] L. von Claisen, P. Roosen, Über die Einwirkung des Phenylhydrazines auf Oxymethylenaceton und auf Acetonoxalsäure, Justus Liebigs Ann. Chem. 278 (1894) 274–296.
- [12] I.L. Finar, G.H. Lord, The formylation of the pyrazole nucleus, J. Chem. Soc. (1957) 3314-3315.
- [13] I.L. Finar, K.E. Godfrey, The preparation and properties of some derivatives of 1-phenylpyrazoles, J. Chem Soc. (1954) 2293–2298.
- [14] C.A. Rojahn, Über die Isomerie-verhältnisse in der Pyrazol-Reihe, Chem. Ber. 59 (1926) 607.
- [15] E. Benary, Zur Kenntnis der Oxymethylen-ketone, insbesondere aus Methyl-äthyl-keton, Chem. Ber. 59 (1926) 2198–2201.
- [16] G. Sorba, A. Di Stilo, C. Medana, C. Cena, A. Gasco, M. Orsetti, The cyano-NNO-azoxy function in the design of an irreversible label for α<sub>1</sub>-adrenoceptor, Bioorg. Med. Chem. 3 (1995) 173–178.
- [17] P.A. Carrupt, P. Gaillard, F. Billois, P. Weber, B. Testa, C. Meyer, S. Pérez, The molecular lipophilicity potential (MLP): a new tool for log P calculation and in comparative molecular field analysis (CoMFA). In: V. Pliska, B. Testa, H. van de Waterbeemd (Eds.), Lipophilicity in Drug Research, VCH, Weinheim, 1996, pp. 195–216.
- [18] Forcefields. In: DISCOVER 95.0/3.0.0 User Guide, Part 1, 3.1-3.33, Molecular Simulations, San Diego, CA.
- [19] M.J.D. Dewar, E.G. Zoebisch, E.F. Healy, J.P. Stewart, AM1: a new general purpose quantum mechanical molecular model, J. Am. Chem. Soc. 107 (1985) 3902–3909.
- [20] S.A. Buckner, K.W. Oheim, P.A. Morse, S.M. Knepper, A.A. Hancock, α<sub>1</sub>-Adrenoceptor-induced contractility in rat aorta is mediated by the α<sub>1D</sub> subtype, Eur. J. Pharmacol. 297 (1996) 241–248.